

S0960-894X(96)00052-2

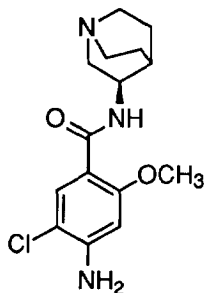
(R)-3-(6-CHLORO-1-ISOPROPYLBENZIMIDAZOLE-4-CARBOXAMIDO)QUINUCLIDINE: A HIGH AFFINITY LIGAND FOR THE (R)-ZACOPRIDE BINDING SITE

L. A. Flippin,^{*} D. S. Carter,¹ J. Berger,¹ R. D. Clark,¹ D. W. Bonhaus,^{*2} E. Leung,² and R. M. Eglén²

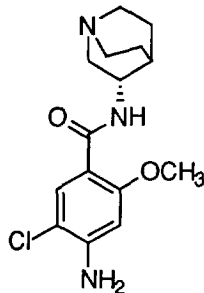
Divisions of Chemical Research and Development and Neurobiology, Roche Bioscience, 3401 Hillview Avenue, Palo Alto, California 94304

Abstract: The (R)-3-amido quinuclidine **6** (RS-16566) was found to be a high affinity ligand for the (R)-zacopride binding site.

Zacopride is a well-known racemic serotonergic agent that exhibits properties of both 5HT₃ antagonism³ and 5HT₄ agonism.⁴ The order of binding affinity of the zacopride antipodes is (S)-zacopride > (R)-zacopride at both of these serotonin receptor subtypes; however, recent reports have described a non-serotonergic binding site in rat cerebral cortex and NG 108-15 clonal cells that exhibits (R)-zacopride > (S)-zacopride selectivity.^{5,6} The extant pharmacology and distribution studies of the so-called (R)-zacopride binding site suggest a novel, high affinity locus with a wide distribution in central and peripheral tissues.^{5b} This binding site has so far been primarily characterized by radioligand binding studies with [³H]-(R)-zacopride; however, emerging in vivo data suggest that it may exhibit a functional role in the behavioral actions of (R)-zacopride that is not simply accounted for by serotonergic properties.⁷

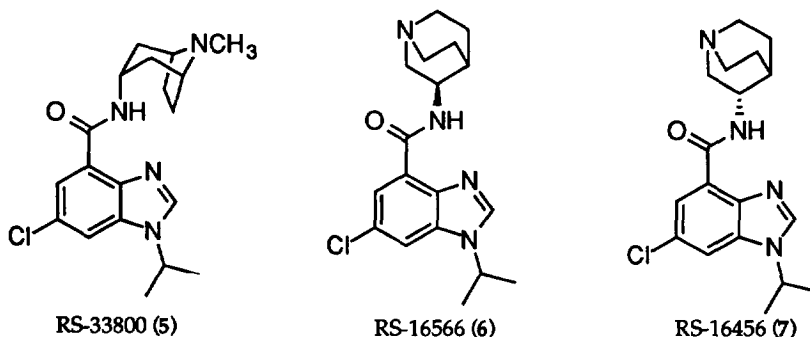


(R)-zacopride



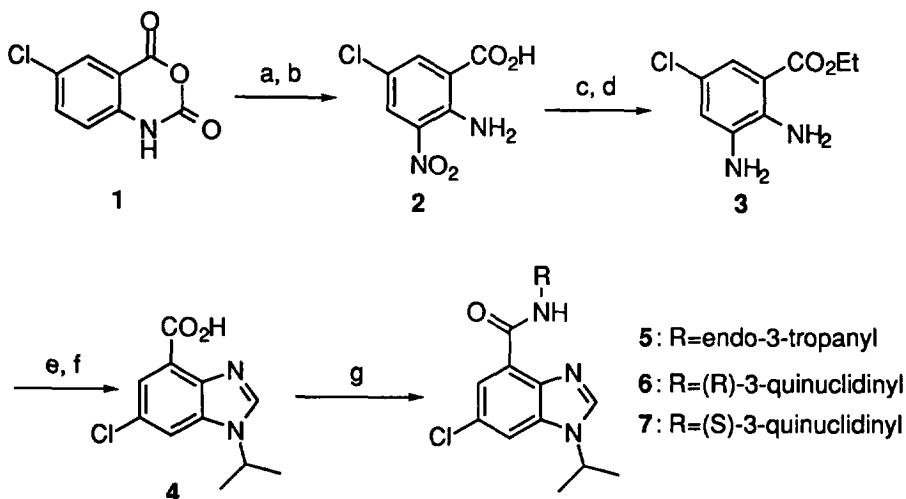
(S)-zacopride

During the course of a program aimed at discovery of 5HT₃ and 5HT₄ receptor ligands we noted that the achiral lead RS-33800, endo-3-(6-chloro-1-isopropylbenzimidazole-4-carboxamido)tropane (**5**), displayed a relatively high affinity (16 nM) for the (R)-zacopride binding site ((R)-ZBS) in NG 108-15 clonal cells. The binding affinity of **5** is roughly comparable to that of (R)-zacopride itself using the NG 108-15 cell line; therefore, we briefly explored the effect of substituting (R)-3-quinuclidinyl and (S)-3-quinuclidinyl moieties for the tropane ring system to give the enantiomeric amides RS-16566 (**6**) and RS-16456 (**7**), respectively.



The 6-chloro-1-isopropylbenzimidazole-4-carboxamides **5-7** were prepared from commercially available 5-chloroisatoic anhydride (**1**) as shown in Scheme I. Nitration of **1**, followed by hydrolysis of the anhydride group, gave 5-chloro-3-nitroanthranilic acid **2** in 80 % yield. Esterification of compound **2** with satd. ethanolic HCl followed by reduction of the nitro group with H_2 -10 % Pd/C afforded ethyl 5-chloro-3-aminoanthranilate **3** in quantitative yield. The 3-amino group was selectively monoalkylated by warming a solution of **3** in 1:1 2-iodopropane-DMF to 50 °C for 6 h; chromatographic purification of the N-monoalkylated product, followed by treatment with formic acid in hot aq. HCl, gave 6-chloro-1-isopropylbenzimidazole-4-carboxylic acid (**4**) in 35 % yield. Carboxylic acid **4** was coupled with endo-3-aminotropane, (R)-3-aminoquinuclidine, or (S)-3-aminoquinuclidine using carbonyl diimidazole in DMF solution to afford amides **5-7** respectively. Compounds **5-7** were converted to their hydrochloride salt forms for all subsequent uses.

Scheme I



a: $NaNO_3/H_2SO_4$ **b**: H_2O , reflux **c**: EtOH-HCl **d**: H_2 -10 % Pd/C **e**: 1:1 DMF-2-iodopropane (50 °C, 6 h) **f**: HCO_2H -aq. HCl, reflux **g**: CDI-DMF; then RNH_2

Selected receptor binding affinities for compounds 5-7 and the zacopride enantiomers are given in the Table.

TABLE. Receptor Binding Affinities of 5-7 and (R)- and (S)-zacopride			
Compound	Binding pK_i^a		
	5HT ₃ ^b	5HT ₄ ^c	(R)-ZBS ^d
5	9.0±0.1	6.80±0.07	7.70±0.10
6	9.27±0.03	7.62±0.01	9.84±0.42
7	9.86±0.08	6.8 ^e	6.22±0.30
(S)-zacopride	9.74±0.03	6.36±0.12	5.3 ^f
(R)-zacopride	8.43 ^g	5.55±0.13	8.30±0.22

^aExcept as noted, radioligand binding assays were carried out as described in reference 8. Values are mean $pK_i \pm \text{SEM}$ ($n \geq 3$).
^bDisplacement of [³H]-BRL 43694 from NG-108-15 cell membranes.
^cDisplacement of [³H]-GR 113808 from Guinea pig brain striatum.
^dDisplacement of [³H]-(R)-zacopride from ondansetron-treated NG-108-15 cell membranes. ^e pK_b estimate from relaxation of carbachol-contracted rat esophageal muscularis mucosae. ^fReference 5.
^gDisplacement of racemic [³H]-zacopride from rat cortex (reference 3b).

All of the compounds in the Table exhibit high affinity for the 5HT₃ receptor and low-to-moderate affinity for the 5HT₄ receptor; however, the range of affinities of these compounds at the (R)-zacopride binding site ((R)-ZBS) is remarkable. Thus, the (R)-antipode of zacopride shows a 1000-fold higher (R)-ZBS affinity than does (S)-zacopride.⁵ (R)-Amido quinuclidine 6, which shows 4000-fold higher affinity at the (R)-zacopride binding site than the corresponding (S)-antipode 7, also exhibits a 35-fold improvement in (R)-ZBS binding affinity over (R)-zacopride itself. Unfortunately, while both (S)-zacopride and compound 7 show excellent 5HT₃/(R)-ZBS selectivity, no ligands are known to date which exhibit useful (R)-ZBS/5HT₃ selectivity. The achievement of *ca.* 100-fold (R)-ZBS/5HT₃ selectivity in a high affinity ligand would clearly provide a powerful pharmacological tool for the further assessment of physiological functions of the (R)-zacopride binding site; thus, additional structure-activity studies are needed toward this end.

References and Notes

^vThis paper is dedicated to Professor Clayton H. Heathcock on the occasion of his 60th birthday.

1. Division of Chemical Research and Development.
2. Division of Neurobiology
3. (a) Pinkus, L. M.; Sarbin, N. S.; Gordon, J. C.; Munson, H. R. *Eur. J. Pharmacol.* **1990**, *179*, 231. (b) Waeber, C.; Pinkus, L. M.; Palacios, J. M. *Eur. J. Pharmacol.* **1990**, *181*, 283.
4. Eglen, R. M.; Swank, S. R.; Walsh, L. K. M.; Whiting, R. L. *Br. J. Pharmacol.* **1990**, *101*, 513.
5. (a) Kidd, E. J.; Bouchelet de Vendgies, I.; Levy, J. C.; Hamon, M.; Gozlan, H. *Eur. J. Pharmacol.* **1992**, *211*, 133. (b) Kidd, E. J.; Levy, J. C.; Nielsen, M.; Hamon, M.; Gozlan, H. *Eur. J. Pharmacol. (Mol. Pharm. Sec.)* **1993**, *247*, 45.
6. Eglen, R. M. *Exp. Opin. Invest. Drugs* **1995**, *4*, 229.
7. (a) Barnes, J. M.; Barnes, N. M.; Costall, B.; Domeny, A.; Johnson, D. N.; Kelly, M. E.; Munson, H. R.; Naylor, R. J.; Young, R. *Pharmacol. Biochem. Behav.* **1990**, *37*, 717. (b) Young, R.; Johnson, D. N. *Eur. J. Pharmacol.* **1991**, *201*, 151. (c) Martin, P.; Gozlan, H.; Peuch, A. *Eur. J. Pharmacol.* **1992**, *212*, 73. (d) Barnes, N. M.; Cheng, C. H. K.; Costall, B.; Ge, J.; Kelly, M. E.; Naylor, R. J. *Eur. J. Pharmacol.* **1992**, *218*, 91.
8. **5HT₃ receptor binding assay:** NG-108 cells were grown in Dulbecco's modified Eagle's medium supplemented with 10 % bovine calf serum and 1 X hypoxanthine-aminopterin-thymidine. Confluent cells were harvested from 250 mL flasks using a 1 min exposure to trypsin. The cell suspension was centrifuged (200 x G; 5 min), resuspended in a Tris (50 mM) EDTA (5 mM) buffer (pH 7.4 @ 4 °C), homogenized using a Polytron P10 tissue disrupter (setting 5 for 10 sec), and recentrifuged for 15 min. For competition binding assays the NG-108 cell membranes were incubated with 1.0 nM [³H]-BRL 43694 and competing ligands in 0.25 mL of Tris-Krebs buffer (154 mM NaCl, 5.4 mM KCl, 1.2 mM K₂PO₄, 2.5 mM CaCl₂, 1 mM MgCl₂, 11 mM D-glucose and 10 mM Tris; pH 7.4 @ 25 °C) for 45 min. Reactions were terminated by vacuum filtration through filters pretreated with 0.3 % PEI. Nonspecific binding was determined with 1 μM BRL 43694. **(R)-zacopride binding site assay:** NG-108 cell membranes, prepared as above, were incubated with 2 nM [³H]-(R)-zacopride and competing ligands in 0.5 mL of Tris buffer (25 mM Tris-HCl; pH 7.4 @ 30 °C). 1 μM Ondansetron was included in the incubation to prevent [³H]-(R)-zacopride binding to 5HT₃ receptors. Reactions were terminated after 30 min by vacuum filtration. Nonspecific binding was defined with 100 μM mianserin. **5HT₄ receptor binding assay:** Guinea pig brain striata were dissected from Guinea pig brains (Rockland Inc., Gilbertsville, PA). Membranes were prepared by homogenization in a Tris buffer (10 mM Tris, 250 mM sucrose, 5 mM EDTA; pH 7.4 @ 4 °C), filtration through nylon mesh, and centrifugation of the filtrate (1000 x G for 15 min). Membranes were incubated with 0.1 nM [³H]-GR 113808 and competing ligands in 0.5 mL of a HEPES buffer (50 mM HEPES, 130 mM choline chloride, 5 mM D-glucose, 5.4 mM KCl, 0.5 mM EDTA; pH 7.4 @ 25 °C). Reactions were terminated after 1 h by vacuum filtration. Nonspecific binding was defined with 1 μM GR 113808.

(Received in USA 23 December 1995; accepted 23 January 1996)